

## The Importance of Differentiating Exposure from Infection with *Borrelia burgdorferi* in the Diagnosis and Treatment of Canine Lyme Disease

The C<sub>6</sub> technology used by IDEXX in USDA-licensed kits differentiates Lyme-infected from vaccinated dogs

### Introduction

The C<sub>6</sub> peptide used in the IDEXX SNAP® 3Dx®, SNAP® 4Dx® Plus and Lyme Quant C<sub>6</sub>® tests has played a foundational role in veterinary infectious disease diagnostics for the last 10 years and has been employed as a method of Lyme disease surveillance in dogs across North America. Recently, the Centers for Disease Control and Prevention (CDC) has recognized that Lyme surveillance data from dogs can be a valuable tool for predicting the emergence of Lyme disease in humans within new geographic regions.<sup>1</sup> This diagnostic update on Lyme C<sub>6</sub> testing will provide information on:

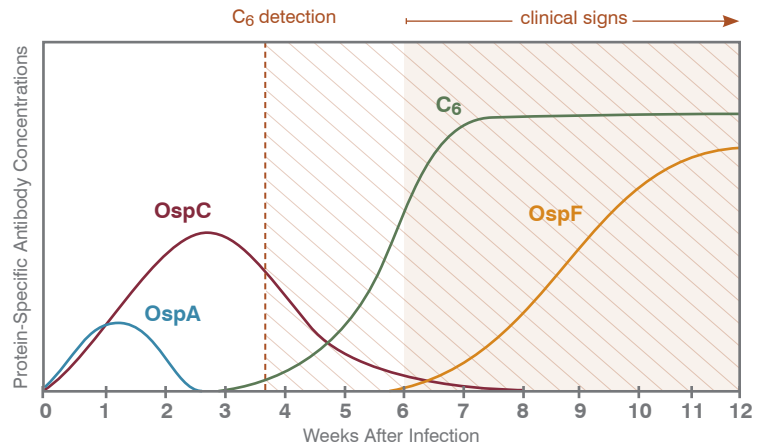
- Why VlsE (C<sub>6</sub>) is expressed by the spirochete.
- How the C<sub>6</sub> peptide is used in Lyme diagnostics.
- When quantitative measures of C<sub>6</sub> may help in the management of subclinical cases of *Borrelia burgdorferi* infections.

### Background

The C<sub>6</sub> peptide represents one of the constant or invariable regions (IR<sub>6</sub>) of the VlsE protein. VlsE is a surface protein of *B. burgdorferi*. It is encoded by the VlsE gene, which contains numerous variable sequences along with the six constant region sequences. The organism selects and expresses different variable sequences over time in order to successfully evade the host immune response and survive within the mammalian host. The VlsE gene is only expressed in the mammalian host. The gene is not expressed when the organism is within the tick or when it is grown in culture to produce the Lyme vaccine. Therefore, antibodies generated as a result of Lyme vaccination do not react with the C<sub>6</sub> peptide.<sup>2</sup> Antibodies to the C<sub>6</sub> peptide are an indication of natural infection with *B. burgdorferi*.

### Surface proteins change with different stages of infection

Osp proteins, like VlsE, are also surface proteins of *B. burgdorferi*. OspA is important for localization of the spirochete within the midgut of the tick and is typically not expressed by the spirochete when it is in the mammalian host. OspC begins to be expressed by the spirochete as the tick takes its blood meal. Expression of OspC facilitates movement of the spirochete from the midgut of the tick to tissues of the host. Both OspA and OspC are targets of Lyme vaccines. OspF is generally expressed 4–6 weeks after the spirochete enters the mammalian host with a resulting antibody response that occurs 6–9 weeks postinfection<sup>3</sup> (see figure 1).



**Figure 1.** Schematic representation of antibody concentrations to different outer surface proteins of *B. burgdorferi*. Antibodies to C<sub>6</sub> precede the onset of clinical signs and indicate infection.

### Host immune response to OspC

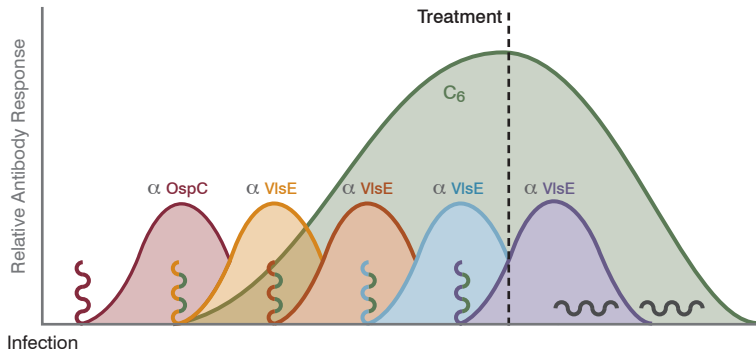
The early host immune response is primarily directed against OspC. As host antibody titers to this protein increase, the spirochete must turn off expression of OspC or risk elimination from the host. OspC expression is greatly reduced by 10 days postinfection,<sup>4–6</sup> and the OspC antibody response may be variable or begin to wane as early as 49 days postinfection.<sup>3,7</sup>

### C<sub>6</sub> antibodies are indicative of infection not exposure

As OspC expression begins to decrease by 10 days postinfection, the VlsE gene is being turned on to allow the organism to employ a novel mechanism of host evasion; it disguises itself with a variable array of different coat proteins. The VlsE protein is only produced after the bacteria has been in the mammalian host for 7–21 days.<sup>4</sup> Antibodies to C<sub>6</sub> indicate infection because the spirochete must infect the dog and be biologically active for at least a week before sufficient amounts of VlsE protein are produced to stimulate the antibody response. Antibodies to C<sub>6</sub> may be detected as early as 3–4 weeks postinfection.<sup>2</sup> Dogs that are protected from infection by vaccination may make more antibodies to OspA and OspC in response to tick-transmitted organisms. However, they do not make antibodies to the C<sub>6</sub> peptide. The organisms appear to be controlled by the immune system prior to the expression of VlsE; this observation provides additional evidence that antibodies to C<sub>6</sub> distinguish infection from exposure.<sup>8</sup>

## Increased levels of C<sub>6</sub> antibodies reflect active infection

Biological activity on the part of the organism is reflected by continuous antigenic variation in response to immune pressure from the host. In other words, an active spirochete must continue to disguise itself with new coat proteins to evade the antibody response of the host. Each new coat (VisE) protein that is expressed also contains C<sub>6</sub>. So, as the dog makes antibodies to the new coat proteins, it also makes more antibodies that react with the C<sub>6</sub> peptide. Experimental infection studies have demonstrated that the higher the C<sub>6</sub> antibody levels, the greater the number of organisms that could be recovered from the skin or tissues of infected dogs. Furthermore, these studies have shown that these organisms are more likely to survive in culture when removed from the dog.<sup>2,9,10</sup> Concentrations of C<sub>6</sub> antibody decline rapidly in response to antibiotic therapy and so do the numbers of organisms that can be recovered from the dog.<sup>2,9,10</sup> As the organisms are eliminated or driven to a state of dormancy, they are no longer changing their coat proteins in an attempt to evade the immune response. There is no further stimulation of the C<sub>6</sub> antibody response and as a result, C<sub>6</sub> antibody concentrations decline (see figure 2).



**Figure 2. Active *B. burgdorferi* infections stimulate the C<sub>6</sub> antibody response.**

At initial infection, a spirochete ( $\beta$ ) expresses OspC on its surface (red). With time, it must change the coat protein to evade the immune response ( $\alpha$  OspC, red line). So by 10 days postinfection, the spirochete no longer expresses OspC and instead begins to express different VisE coat proteins over time, each of which includes C<sub>6</sub>. This drives C<sub>6</sub> antibody concentrations higher. Following treatment, the organism becomes dormant, no longer expresses VisE and C<sub>6</sub> antibody concentrations decline.

## Quantitative concentrations of C<sub>6</sub> antibodies help to identify and monitor dogs that may benefit from treatment.

Because quantitative C<sub>6</sub> antibody concentrations correlate with organism load and viability, the Lyme Quant C<sub>6</sub><sup>®</sup> Test can help to identify *B. burgdorferi*-infected dogs that would benefit from antibiotic therapy, including those that lack the more recognizable clinical signs or laboratory abnormalities of Lyme disease. In general, *B. burgdorferi*-infected dogs, humans and nonhuman primates with high concentrations of C<sub>6</sub> antibody respond to antibiotic therapy with a marked reduction in C<sub>6</sub> antibody concentrations.<sup>9,11,12</sup> Thus, even in dogs that show no clinical signs, treatment response can be monitored by measuring a reduction in the concentrations of C<sub>6</sub> antibody. Failure of the test to show a reduction in the concentration of C<sub>6</sub> antibody following treatment

may indicate treatment failure, recrudescence, noncompliance with administering medication or reexposure to *B. burgdorferi*. C<sub>6</sub> antibody concentrations do not correlate with disease or predict which dogs will become sick with Lyme disease or Lyme nephritis. Dogs with lower concentrations of C<sub>6</sub> antibodies (<30 U/mL) may or may not benefit from antibiotic treatment; a means to monitor response to therapy in this population is currently not available.

## Clinical uses and advantages of C<sub>6</sub> testing

Testing for antibodies to C<sub>6</sub> as part of the vector-borne disease screening protocol is easy with the IDEXX SNAP<sup>®</sup> 3Dx<sup>®</sup>, SNAP<sup>®</sup> 4Dx<sup>®</sup> Plus and Lyme Quant C<sub>6</sub><sup>®</sup> tests. It provides a number of unique benefits for veterinarians:

- Most important, antibodies to C<sub>6</sub> are specific for infection, which minimizes the risk of false positives and avoids confusion with Lyme-vaccinated patients.
- C<sub>6</sub> also distinguishes between infection and exposure, so practitioners can be confident in their diagnostic conclusions and effectively recommend the appropriate diagnostic tests and treatment for each patient.
- Furthermore, with regard to therapy, C<sub>6</sub> provides an effective means to monitor treatment, as declining concentrations of C<sub>6</sub> antibody indicate a reduction in organism load and viability.

## References

1. Mead P, Goel R, Kugeler K. Canine serology as adjunct to human Lyme disease surveillance. *Emerg Infect Dis.* 2011;17(9):1710–1712.
2. Liang FT, Jacobson RH, Straubinger RK, Grooters A, Philipp MT. Characterization of a *Borrelia burgdorferi* VisE invariable region useful in canine Lyme disease serodiagnosis by enzyme-linked immunosorbent assay. *J Clin Microbiol.* 2000;38(11):4160–4166.
3. Wagner B, Freer H, Rollins A, et al. Antibodies to *Borrelia burgdorferi* OspA, OspC, OspF, and C6 antigens as markers for early and late infection in dogs. *Clin Vaccine Immunol.* 2012;19(4):527–535.
4. Crother TR, Champion CI, Whitelegge JP, et al. Temporal analysis of the antigenic composition of *Borrelia burgdorferi* during infection in rabbit skin. *Infect Immun.* 2004;72(9):5063–5072.
5. Liang FT, Jacobs MB, Bowers LC, Philipp MT. An immune evasion mechanism for spirochetal persistence in Lyme borreliosis. *J Exp Med.* 2002;195(4):415–422.
6. Liang FT, Nelson FK, Fikrig E. Molecular adaptation of *Borrelia burgdorferi* in the murine host. *J Exp Med.* 2002;196(2):275–280.
7. Magnarelli LA, Levy SA, Ijdo JW, Wu C, Padula SJ, Fikrig E. Reactivity of dog sera to whole-cell or recombinant antigens of *Borrelia burgdorferi* by ELISA and immunoblot analysis. *J Med Microbiol.* 2001;50(10):889–895.
8. LaFleur RL, Callister SM, Dant JC, et al. One-year duration of immunity induced by vaccination with a canine Lyme disease bacterin. *Clin Vaccine Immunol.* 2010;17(5):870–874.
9. Philipp MT, Bowers LC, Fawcett PT, et al. Antibody response to Ig<sub>G</sub>, a conserved immunodominant region of the VisE lipoprotein, wanes rapidly after antibiotic treatment of *Borrelia burgdorferi* infection in experimental animals and in humans. *J Infect Dis.* 2001;184(7):870–878.
10. Straubinger RK. PCR-based quantification of *Borrelia burgdorferi* organisms in canine tissues over a 500-day postinfection period. *J Clin Microbiol.* 2000;38(6):2191–2199.
11. Levy SA, O'Connor TP, Hanscom JL, Shields P, Lorentzen L, Dimarco AA. Quantitative measurement of C<sub>6</sub> antibody following antibiotic treatment of *Borrelia burgdorferi* antibody-positive nonclinical dogs. *Clin Vaccine Immunol.* 2008;15(1):115–119.
12. Embers ME, Barthold SW, Borda JT, et al. Persistence of *Borrelia burgdorferi* in rhesus macaques following antibiotic treatment of disseminated infection. *PLOS ONE.* 2012;7(1):e29914. [www.plosone.org/article/info:doi/10.1371/journal.pone.0029914](http://www.plosone.org/article/info:doi/10.1371/journal.pone.0029914). Accessed October 12, 2012.